

Assessment of diagnostic methods for urinary schistosomiasis, Assalya, White Nile State, Sudan

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Abstract:

Background: *Schistosomiasis is a parasitic disease caused by blood flukes (trematodes) of the genus Schistosoma. It is the third most devastating tropical disease in the world, being a major source of morbidity and mortality for developing countries. The study aim to compare macrohaematuria and urine dipstick against intensity of urinary schistosomiasis and calculate sensitivity and specificity of macrohaematuria and urine dipstick in samples collected from children who attended Assalaya medical center.*

Methods: *This cross sectional study conducted in Assalaya, White Nile State, Sudan. Four hundred and twenty participants were included, 246 (58.6%) were males and 174 (41.4%) were females. The*

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age ranged between (9-17 years). Urine samples were collected and examined for Schistosoma haematobium by using macrohaematuria, dipsticks for proteinuria and microhaematuria, filtration method as gold standard method.

Results: *Macrohaematuria, microhaematuria and proteinuria showed a sensitivity of 100%, 100% and 100% with specificity of 19.8%, 14.56% and 9.84% respectively. Association of intensity of infection with macrohaematuria, microhaematuria and proteinuria was significant. $P < 0.05$.*

Conclusion: *The study concluded that urine dipstick strip and macrohaematuria are highly sensitive with low specificity in detection of S. haematobium in urine samples. In addition, they have potential value to reflect the intensity of infection.*

Key words: urinary schistosomiasis - urine dipstick strips -Sudan

INTRODUCTION

Schistosomiasis is a parasitic disease caused by blood flukes (trematodes) of the genus *Schistosoma*. After malaria and intestinal helminthiasis, schistosomiasis is the third most devastating tropical disease in the world, being a major source of morbidity and mortality for developing countries in Africa.⁽¹⁾ *S. haematobium* affects both the urinary and reproductive tract systems.⁽²⁾ *S. haematobium* infection causes haematuria, dysuria, lesions of the bladder, kidney failure, bladder cancer.⁽³⁾ Infection also interferes with nutrient uptake and can lead to undernutrition, growth and cognitive development retardation, and pose a serious threat to children's health, education and productivity.⁽⁴⁾ As the transmission of many other parasitic diseases, the transmission of schistosomiasis is a complex process governed by natural, socio-economic factors and human life style.^(5, 6) Disease transmission occurs in ecosystems that support both the snails that serve as the obligate intermediate host for the parasite and the parasite itself. Humans become

infect through contaminated freshwater when free-swimming cercariae, which are released from the snail host, penetrate the skin. Humans must have contact with contaminated freshwater in order to acquire *Schistosoma* infection.⁽⁶⁾ The prevalence of schistosomiasis increases when new facilities for either irrigation canals or construction of dams create the vector (snail).⁽⁷⁾ Introduced as early as 1909 in Dongola and Southeastern Sudan. The highest infectivity was reached following the implementation of Gezira cotton irrigation scheme.⁽⁸⁾ The objective of this study is to compare the intensity as measured through macrohaematuria and urine dipstick and evaluate the sensitivity and specificity of the two techniques in samples collected from children who attended Assalaya medical Centre.

MATERIALS AND METHODS

Type of study

Descriptive cross sectional study.

Study area

Assalaya campus is located at Assalaya administrative unit, Rabak Locality, White Nile State. The source of water supply for sugar cane farms is White Nile from which canals enable irrigation. The main occupation of the population is farming in Sugar Cane farms.

Study population

School age children referred to Assalaya, White Nile State medical Centre.

Study period

The study commenced in June and ended in October 2014

Sample collection

Four hundred and twenty urine samples collected in clean and labelled containers. Minimum of fifteen ml of urine collected for each sample.

METHODOLOGY

Macrohaematuria

Macroscopic examination done for detection of visible haematuria (macrohaematuria) by naked eye

Diagnostic dipstick strip method:

Microhaematuria

In the laboratory, a reagent strip (*Mission® Expert Urinalysis Strips*) carefully dipped into urine sample and allowed to stand for 5seconds. Moreover, the colour matched with the standard colour by the side of the container as recommended by the manufacturer to estimate the amount of blood in the urine

Urine Protein Analysis (proteinuria)

The protein excretion in the urine was determined using simple reagent strips (*Mission® Expert Urinalysis Strips*). Urinary protein results categorized as positive or negative according to the manufacturer's instruction provided in the kit inserts.

Filtration method

Urine samples examined with the urine filtration method for the presence of *S. haematobium* eggs. In brief, urine samples were vigorously shaken, and 10 ml of each sample were pressed through a 13-mm diameter small-meshed filter (20 µm pores diameter) for detection of *S. haematobium* eggs, and infections categorised as light (≤ 50 eggs) and heavy (≥ 50 eggs). Each filter placed on a microscope slide labelled with the child's identifier

Statistical Analysis

The data from the findings were analyzed using SPSS statistic version 20. Relationship of proteinuria, microhaematuria and macrohaematuria were tested using Chi square and Fisher's Exact test. P-value < 0.05 considered significant. The results of the analysis presented in figures and tables

The sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV) was calculated using filtration method as the gold standard.

RESULTS

Total 420 children 246 (58.6%) were males 210 (85%) of these were infected and 174 (41.4%) were females 97(56%) of these were infected. The age ranged between (9-17 years). Different diagnostic methods used to detect intensity of *S. haematobium*. Filtration method used to assess the intensity of infection and categorized into two groups, heavy (egg count 50 and more eggs/10 ml) and light (egg count less than 50 eggs/10 ml), while macrohaematuria, microhaematuria and proteinuria recorded as Present or absent.

Table 1: Distribution of age groups and sex

Variables	Total	
Sex	Males	246
	Females	174
Age	9-11	184
	12-14	172
	15-17	64

Table 2: Shows association of intensity of *S. haematobium* with Macrohaematuria, microhaematuria and proteinuria

		<i>Intensity of infection</i>			<i>P. value</i>
		Light	Heavy	Total	
<i>Macrohaematuria</i>	Present	81	98	179	0.000
	Absent	20	0	20	
	Total	101	98	199	
<i>Microhaematuria</i>	Present	88	98	186	0.000
	Absent	15	0	15	
	Total	103	98	201	
<i>Proteinuria</i>	Present	110	71	181	0.003
	Absent	12	0	12	
	Total	122	71	193	

Table 3: Shows sensitivity and Specificity of microhaematuria, macrohaematuria and proteinuria

	SENSITIVITY (%)	SPECIFICITY (%)	PPV	NPV
PROTEINURIA	100	9.84	56	87
MACROHAEMATURIA	100	19.80	65	86
MICROHAEMATURIA	100	14.56	69	90

DISCUSSION

The use of macrohaematuria, microhaematuria and proteinuria to estimate the intensity of urinary schistosomiasis has potential to give a clue about intensity of infection among infected children.

The association between macrohaematuria, microhaematuria and proteinuria with intensity of infection was significant.

Macrohaematuria in this study, increased with intensity of *S. haematobium* infection. As *Akyala Ishaku* represented in his study in *Nigeria* ⁽⁹⁾ and *Ugbomoiko et al* ⁽¹⁰⁾, the presence of macrohaematuria use as prove for infection it can be a sign of tissue damage due to infection and agree with *Bogoch et al* ⁽¹¹⁾ in northern Ghana, self-reporting haematuria (macrohaematuria) increased with intensity of infection.

Study in Mali, *Moussa Sacko* ⁽¹²⁾ reported haematuria related to intensity of infection and microhaematuria strongly related to number of *S.haematobium* eggs. The results indicate that blood in urine provides a good sound of urinary schistosomiasis. *Ugbomoiko et al* ⁽¹⁰⁾ confirm that any patients with heavy infection, microhaematuria detected, hence microhaematuria increased with severity of infection.

The significant association between proteinuria and intensity of infection in our study similar to that finding by *Bogoch et al* ⁽¹¹⁾ in northern Ghana.

In this study, showed macrohaematuria and dipstick high sensitive and low specific to *S.haematobium* infection, high sensitivity of parameters in this result may due to other disease in urinary tract, condensation of urine, which appear dark, occult blood and sensitivity of dipstick itself but not related to *S.haematobium* infection. In the other hand children more susceptible to nutritional defect lead to presence of protein in urine in high amount, in contrast to study of *Ugbomoiko* ⁽¹⁰⁾ *Koukounari et al.*⁽¹³⁾ who found haematuria dipsticks low sensitive and high specific, while specificity was high in spite of low sensitivity. *Ugbomoiko* observed sensitivity increase with intensity of infection and obviate that by combinations of them. *Bogoch et al* ⁽¹¹⁾ found macrohaematuria and proteinuria are high specificity with low sensitivity, while

microhaematuria high sensitive and specific which indicate high amount of eggs in patients.

Morenikeji (14) macrohaematuria, microhaematuria and proteinuria were low sensitivity and high specificity and *Mtasiwa D* (15) findings low sensitivity and high specificity of microhaematuria and macrohaematuria which mean morbidity in urinary tract due to *S. haematobium*. *Houmsou* (16) The sensitivity of urine colour for screening test low and high specificity which give better indicator to damage or complication occur in renal and urinary tract .

Our finding contrast with study carried in the Egypt among schoolchildren, by *Kotb MM* (17) who found that the study revealed that microhaematuria with sensitive and proteinuria with sensitivity low and specificity high. In another study carried by *Ndyomug* in Dar-es-Salaam, Tanzania (18), detected microhaematuria sensitivity lower than the sensitivity of microhaematuria detected in this study.

The use of urine dipstick strips has good offer as an indirect method in diagnosis of *S. haematobium* infection.

CONCLUSION

Urine dipstick strip and macrohaematuria are highly sensitive with low specificity in detection of *S. haematobium* in urine samples. In addition, they have potential value to reflect the intensity of infection.

RECOMMENDATIONS:

- 1- Using filtration method (gold standard) to detect intensity of *S. haematobium*.
- 2- Using macrohaematuria, microhaematuria and proteinuria urine dipstick strips is practical-cheap, fast

and easy to use in estimating intensity of *S. haematobium* infection.

- 3- Annual examination of school children usually conduct by the ministry of health should continue.
- 4- School children should be given special consideration in the health education programme, as they are always ready to listen and learn compared to other age groups.
- 5- Public education is very important as to the use of their home latrines and to avoid contact with canal water by constructing bridges across the canal.

REFERENCES:

1. Adisa J, Egbujo EM, Yahaya BA, Echejoh G. Primary infertility associated with schistosoma mansoni: a case report from the Jos plateau, north central Nigeria. African Health Sciences. 2012;12(4):563-5.
2. Nour NM. Schistosomiasis: Health Effects on Women. Reviews in Obstetrics and Gynecology. 2010;3(1):28-32.
3. Fenwick A, Savioli L, Engels D, Robert Bergquist N, Todd MH. Drugs for the control of parasitic diseases: current status and development in schistosomiasis. Trends in parasitology. 2003;19(11):509-15.
4. Deribew K, Tekeste Z, Petros B, Huat LB. Urinary schistosomiasis and malaria associated anemia in Ethiopia. Asian Pacific journal of tropical biomedicine. 2013;3(4):307-10.
5. Hu H, Gong P, Xu B. Spatially explicit agent-based modelling for schistosomiasis transmission: human-environment interaction simulation and control strategy assessment. Epidemics. 2010;2(2):49-65.
6. Woodhall DM, Wiegand RE, Wellman M, Matey E, Abudho B, Karanja DMS, et al. Use of Geospatial Modeling to

Predict *Schistosoma mansoni* Prevalence in Nyanza Province, Kenya. PLoS ONE. 2013;8(8):e71635.

7. Tayrab E, Ashmaig A, Shareef H, Bedawi S, Aradaib I. Association of *Schistosoma mansoni* with infertility in a Sudanese patient from schistosomiasis area of endemicity: a case report. Research Journal of Medical Sciences. 2010;4(3):125-7.

8. Humaida S, Ahmed A, Homeida MM. Schistosomiasis: epidemiology and burden of disease in the Sudan. Associate Editor. 2011;47(2).

9. Akyala Ishaku. A AD, Tanimu Habibu , Tsaku Mary , Agieni Ashem Godwin , "Microhaematuria and Proteinuria Performance as a Measured by Urine Reagent Strips in Estimating Intensity and Prevalence of Schistosoma haematobium Infection in Nigeria", American Journal of Medicine and Medical Sciences, Vol. 2 No. 6, 2012, pp. 144-148. doi: 10.5923/j.ajmms.20120206.06. Houmsou, RS, Kela S, Suleiman M. Performance of microhaematuria and proteinuria as measured by urine reagent strips in estimating intensity and prevalence of *Schistosoma haematobium* infection in Nigeria 2011.

10. Ugbomoiko US, Dalumo V, Ariza L, Bezerra FS, Heukelbach J. A simple approach improving the performance of urine reagent strips for rapid diagnosis of urinary schistosomiasis in Nigerian schoolchildren. Memorias do Instituto Oswaldo Cruz. 2009;104(3):456-61.

11. Bogoch, II, Andrews JR, Dadzie Ephraim RK, Utzinger J. Simple questionnaire and urine reagent strips compared to microscopy for the diagnosis of *Schistosoma haematobium* in a community in northern Ghana. Tropical medicine & international health : TM & IH. 2012;17(10):1217-21.

12. Sacko M, Magnussen P, Keita AD, Traore MS, Landoure A, Doucoure A, et al. Impact of *Schistosoma haematobium* infection on urinary tract pathology, nutritional status and

anaemia in school-aged children in two different endemic areas of the Niger River Basin, Mali. *Acta tropica*. 2011;120 Suppl 1:S142-50.

13. Koukounari A, Webster JP, Donnelly CA, Bray BC, Naples J, Bosompem K, et al. Sensitivities and specificities of diagnostic tests and infection prevalence of *Schistosoma haematobium* estimated from data on adults in villages northwest of Accra, Ghana. *The American journal of tropical medicine and hygiene*. 2009;80(3):435-41.

14. Morenikeji O, Quazim J, Omoregie C, Hassan A, Nwuba R, Anumudu C, et al. A cross-sectional study on urogenital schistosomiasis in children; haematuria and proteinuria as diagnostic indicators in an endemic rural area of Nigeria. *Afr Health Sci*. 2014;14(2):390-6.

15. Mtasiwa D, Mayombana C, Kilima P, Tanner M. Validation of reagent sticks in diagnosing urinary schistosomiasis in an urban setting. *East African medical journal*. 1996;73(3):198-200.

16. Houmsou R, Amuta EU, Omudu E. The Merits of Urine Color Observation as a Rapid Diagnostic Technique to Estimate *Schistosoma Haematobium* Infection in Two Endemic Areas of Benue State, Nigeria 2012.

17. Kotb MM, Shouman AE, Hussein HM, Khela AK, Kandil SK. Evaluation of the effectiveness of dipstick haematuria and proteinuria in screening *Schistosoma haematobium* infection among school children in upper Egypt. *The Journal of the Egyptian Public Health Association*. 1996;71(5-6):353-67.

18. Ndyomugenyi R, Minjas JN. Urinary schistosomiasis in schoolchildren in Dar-es-Salaam, Tanzania, and the factors influencing its transmission. *Annals of tropical medicine and parasitology*. 2001;95(7):697-706.