

Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease

SANAULLAH KHAN

KALEEMULLAH MANDOKHAIL

HABIB UR REHMAN

Department of Microbiology
University of Balochistan, Quetta, Pakistan

Abstract

The etiology of Type 2 Diabetes Mellitus (T2DM) and Parkinson's disease (PD) is very different, however these two diseases are linked with each other as the Patients suffering from diabetes have 40% more risk for getting Parkinson's disease as compared to diabetes free individuals. In this review article, the dysregulation of some important proteins has been revealed, having distinct association in the pathophysiological mechanisms of T2DM and PD such as protein misfolding and aggregation, autophagy dysregulation, mitochondrial damage (dysfunction), Endoplasmic reticulum stress (malfunction), inflammation and loss of central and peripheral synapse. The proteins involved in the pathogenesis of T2DM and PD through different pathophysiological pathways are amyloid fibres, Parkin & PINK1, Beclin, ATG7, LC3-II, Carnosinase (CNDP2), Synaptophysin, SNAP-25, Sorcin, GLP-1 analogues, AMPK and ADH1A1. Dysregulation of these proteins leads to the pathogenesis of both diseases. In conclusion, the current study suggests that type 2 diabetes mellitus and Parkinson's disease are associated through number of common proteins and Pathophysiological pathways but it is too earlier to conclude that T2DM is a risk factor for the progression of PD. Further studies are required to reveal the relationship of T2DM with PD.

Key words: Type 2 Diabetes Mellitus, Parkinson's disease, Substantia nigra and Proteomics.

1. INTRODUCTION

1.1 Type 2 Diabetes Mellitus (T2DM)

Type 2 Diabetes Mellitus (T2DM) is a complicated and developing disorder characterized by diverse metabolic deficiencies, affecting many organs (Wu et al., 2014). T2DM is caused by combination of genetic, environmental and behavioral factors (Chen et al., 2011; Diabetes mellitus interagency coordinating committee [DMICC], 2011). The lifestyle factors identified to develop T2DM are lack of exercise, inactive lifestyle, smoking and excessive alcohol drinking (Hu et al., 2001). The contribution of obesity in T2DM development is about 90% (World health organization [WHO], 2011). The main factors involved in the development of T2DM are less insulin secretion and resistance of insulin in peripheral tissues like adipose tissues, muscles and liver (Butler et al., 2003; Kahn, 2003; Pratley & Weyer, 2001; Stumvoll et al., 2005). The most common Type of diabetes i.e., T2DM is marked by Hyperglycemia, resistance of insulin and deficiency of insulin (Maitra & Abbas, 2005). New studies evaluated that reduction in the function of α -cell also contributes in the development of T2DM (Fujioka, 2007). The worldwide prevalence of Diabetes mellitus was over 463 million in 2019, in which more than 90% of diabetes patients were diagnosed with T2DM. This prevalence is projected to be 578 million in 2030 and 700 million in 2045 (Saeedi et al., 2019).

1.2 Parkinson's Disease (PD)

Parkinson's disease (PD) is a developing and neurodegenerative disease that influences the regulation of body movements in a person by affecting coordination between brain and other parts of the body for the movements of the muscles (Rewar, 2015). PD usually arises about 60 year's age, but it may emerge earlier (Edinburg Regional Medical Center [ER], 2015). This disorder is developed by degeneration of neurons found in the "Substantia Nigra (SN)" of mid brain and the function of these nerve cells is production of dopamine (Parkinson's disease Information [PDI], 2015; Kumar et al., 2010). The exact cause of PD is still unknown, however both genetic and environmental factors are vital for its exposure (Shafique et al., 2011). The four major symptoms of PD are Tremor, bradykinesia (Slowness

of movement), rigidity (Muscles stiffness), and Postural Instability (Rewar, 2015). Other Symptoms include Dyskinesia, Dementia (Imtiaz et al., 2016), Anosmia (absence of sense of smell), Anxiety, Constipation, Depression, Fatigue, festination of speech, Postural hypotension and Micrographia (Kalia & Lang, 2015; Chao et al., 2015). The worldwide prevalence of PD is about 6.5 million and it is predicted to be double in the upcoming 10-20 years (Imtiaz et al., 2016).

2. PROTEOMICS CORRELATION BETWEEN T2DM AND PD

Type 2 Diabetes mellitus and Parkinson's disease are very different in their etiology; however these two diseases are linked with each other as the Patients suffering from diabetes have 40% more risk for getting Parkinson's disease as compared to diabetes free individuals (Lima et al., 2014; Xu et al., 2011). Several cross-sectional and prospective studies have also revealed the linkages between Type 2 DM and PD through different surveys (Pressley et al., 2003). These studies raised number of questions about the interrelation between these two disorders. Few common molecular mechanisms are also involved among Type 2 DM and Parkinson's diseases including the protein misfolding, aggregation and amyloid synthesis, high secretion of methylglyoxal (MG) and insufficiency of dopamine (Aviles-Olmos et al., 2012; Fatima et al., 2014; Hipkiss, 2012; Tian et al., 2015).

The determination of the linkages between Type 2 Diabetes Mellitus (T2DM) and Parkinson's disease (PD) is very important; involving the common regulation of the proteins in T2DM and Parkinson's disease individuals possibly will reveal the association between these two diseases.

In this review article, the dysregulation of some important proteins has been revealed, having distinct association in the pathophysiological mechanisms of T2DM and PD such as autophagy dysregulation, mitochondrial damage (dysfunction), Endoplasmic reticulum (ER) stress (malfunction), inflammation and loss of central and peripheral synapse.

2.1 Amyloid Fibres

The aggregation of amyloid fibres from unfolded polypeptide plays a vital role in both Parkinson's disease and Type 2 diabetes mellitus, because amyloid structures are synthesized from amylin [islet amyloid polypeptide (IAPP)] in T2DM while amyloid like structures are synthesized from α -synuclein in Parkinson's disease. This linkage of IAPP and α -synuclein in the synthesis of amyloid make the association of these two disorders more complex (Surguchov, 2016).

2.2 Parkin and PINK 1:

Mitophagy is a quality control process in healthy neurons for removal of damaged or non-functional mitochondria to prevent cell death, neuronal cell death is caused by increased mitochondrial damage that leads to the pathogenesis of PD. Mitophagy or the capability of neurons to remove dysfunctional organelles (Mitochondria) is reduced by loss of Parkin or PINK1 (PTEN-induced putative kinase 1) proteins as a result these dysfunctional organelles are accumulated in the neurons and cause the early-onset PD (Pickrell & Youle, 2015). PINK1 identify dysfunction of mitochondria and then signals Parkin to ubiquitinate distinctively the non-functional mitochondria to initiate their removal by autophagy (Pickrell & Youle, 2015). This indicates that PINK1 and Parkin collectively have a mitochondrial quality control role and prevents Parkinsonism in human beings (Geisler et al., 2010). Autophagy has very distinctive role in the homeostasis of islet and improvement of β cell mass against a fat-enriched diet (Marrif & Al-Sunousi, 2016). Certain studies have concluded that the pathway of PINK1/PARKIN perform vital role in mitochondrial quality control in obese and T2DM. PINK1/PARKIN pathway mediated mitochondrial quality control mechanism is reduced by suppressing the transcription of PINK1 level in skeletal muscle tissues of obese and T2DM patients (Scheele et al., 2007). Additionally, pancreatic β cells may be damaged by defective function of PARKIN causing decreased secretion of insulin (Hoshino et al., 2014).

2.3 Beclin:

Beclin is a protein associated with the nucleation phase of autophagy; its level was examined scarcely high in cerebral cortex and

hippocampus of type 2 diabetic mice. The level of β -cell lymphoma 2 (a beclin repressor) was slightly down in the hippocampus of diabetic type 2 mice while the level of PI3K class III kinase (an activator for beclin-induced nucleation) has been examined high in the region of cerebral cortex and hippocampus of type 2 diabetic mice (Carvalho et al., 2015). Autophagy is a type of catabolism involved in the digestion of long-lived proteins and non-functional organelles in the cells of eukaryotes activated by various adverse conditions such as decreased level of nutrients, hypoxia and low energy supply, as a result these catabolic products especially amino acids are released into the cytoplasm for essential biosynthetic mechanisms (Lynch-Day et al., 2012). The primary function of autophagy is protection by regulating nutrient and energy homeostasis in a stressed condition, however it may also be involved in the pathogenesis of many neurodegenerative disorders, such as Alzheimer, Huntington's, and Parkinson's diseases because abnormal proteins and non-functional/damaged organelles are accumulated as a result of dysregulation of autophagy (Banerjee et al., 2010). Dysregulation of autophagy may be caused by abnormal Beclin-1 protein levels contributing in the pathogenesis of neurodegenerative disorders (Pickford et al., 2008; Nascimento-Ferreira et al., 2013; Lucin et al., 2013). For example, the level of Beclin-1 is low in many neurodegenerative diseases (Pickford et al., 2008) while high regulation of Beclin-1 improves the pathogenesis of neurodegenerative diseases in animal models (Nascimento-Ferreira et al., 2013). Autophagy is commonly activated via mTOR (mechanistic target of rapamycin) signaling pathway however additional or secondary signaling pathway for activation of autophagy is the Vps34-Beclin-1 complex to enhance cell survival. For instance, the aggregation of α -synuclein is reduced in case of high regulation of Beclin-1 to minimize cell death and maximize the activity of autophagy (Spencer et al. 2009).

2.4 ATG7:

The main role of ATG (autophagy related proteins) proteins is to control the formation of autophagosome, transportation to lysosomal portion and cargo gathering (Arroyo et al., 2014). Modification in these proteins can accommodate the clearance of impaired organelles and accumulated proteins. The level of ATG7 (Autophagy related 7)

was revealed extremely low in the cerebral cortex and hippocampus regions of type 2 Diabetic mice model, indicating that the phase of elongation for autophagosome synthesis has been agitated (Carvalho et al., 2015). ATG7 is an important autophagy protein needed for membrane transportation and degeneration of axonal terminals, and axonopathy linked with neurodegeneration caused by deterioration of axonal autophagy (Komatsu et al., 2007). Vacuoles resembling autophagosome have also been revealed in the non-functional or degenerating axons linked with various kinds of chronic neurodegenerative disorders which include (Nixon et al., 2005; Cataldo et al., 1996), Parkinson's disease (PD) (Anglade et al., 1997), and animal models for neurodegenerative diseases (Yu et al., 2005; Lin et al., 2003; Li et al., 2001). New research study suggests that minimal levels of autophagy preserve neurodegeneration because autophagy is important for the regulation of local homeostasis of axon terminals and prevent degeneration of axons (Komatsu et al., 2007). These findings indicate an association between locally modified autophagy and axonopathy, which is involved in neurodegeneration (Coleman, 2002).

2.5 LC3-II:

Autophagy is the main process for the breakdown and recycling of prolonged proteins and dysfunctional organelles inside cells (Klionsky, 2000; Lynch-Day et al., 2012). LC3s (MAP1-LC3s) are autophagosomal membranous structural proteins broadly used as biomarkers of autophagy (He et al., 2003; Bai, Inoue et al., 2012). LC3-II (microtubule-associated protein 1A/1B-light chain 3- II) is a type of LC3 which is the major constituent of the autophagosomal membrane found both on the inner and outer surface of the membrane. It has been obtained from a protein known as pro LC3 (30 KDa), which is cleaved by autophagin ATG4 to synthesize the form LC3-I activated by ATG7, and then shifted to ATG3 to develop into LC3- II bounded to the membrane (Ichimura et al., 2000). The LC3-II found in the outer region is discharged to the cytosol and the LC3-II found in the inner region is hydrolyzed by hydrolases after the synthesis of autophagosome (Kabeya et al., 2000). This latest form of LC3-II found on the membranes of autophagosome and autolysosome act as an appropriate marker of autophagy (Kabeya et al., 2000; Wu et

al., 2006). The level of LC3-II protein has been revealed slightly decrease in the cerebral cortex of type 2 Diabetic mice model (Carvalho et al., 2015).

2.6 Carnosinase (CNDP2):

Carnosinases are Xaa-His dipeptidases that have variety of roles in organisms. The isoforms of human carnosinase (CN1 and CN2) speed up the hydrolysis of the dipeptides carnosine (β -alanyl-L-histidine) and homocarnosine (γ -aminobutyryl-L-histidine) under suitable conditions. The deregulation of expression and activity of carnosinase leads to certain physiological defects and disorders like diabetes mellitus, ischemia and neurological disorders (Bellia et al., 2014). Carnosinase, CNDP2 (Carnosine Dipeptidase 2) has shown increase activity in the SN of PD patients through several proteomic studies therefore it is suggested that carnosine (CAR) make effort for protection against PD outbreak by reacting with MG and by interfering with glycolysis (Hipkiss, 2012). Carnosinase influences the metabolism of glucose and prevents diabetic deterioration (Lee et al., 2005; Sauerhofer et al., 2007). It is suggested that the existence of CAR, homocarnosine and anserine in central nervous system and their aging alterations (Hipkiss, 2005; Huang et al., 2005) have a therapeutic characteristic in neurodegenerative diseases (Hipkiss, 2007). Carnosine is neuro-protective because it has the potential to neutralize both oxidative (La-Mendola et al, 2002) and nitrosative stress (Fontana, Pinnen, Lucente & Pecci, 2002) in pathological conditions (Dukic-Stefanovic et al., 2001; Pubill et al., 2002) , like ischemia (Tang et al., 2007). CNDP1 (Carnosine Dipeptidase 1) has low activity levels in neurological disorders including Parkinson's disease (Butterworth et al., 1996). On the other side, CNDP2 is highly expressed in the SN of PD (Licker et al., 2012). Therefore carnosine has a neuroprotective role by lowering neurotoxicity through its antioxidant potential (Trombley et al., 2000). The linkage between normal kidney function and tissue carnosinase activities has been revealed suggesting that CAR may have protective role in human diabetic kidney disease (Janssen et al., 2005). It is also suggested that increased diabetic nephropathy may be due to MG reveals that high level of carnosine may have role in the suppression of MG-mediated pathology. Despite, the observations that pathology is elevated when

the activity of carnosinase is high in both kidney and SN is consistent with the proposal that carnosine may employ some protective activity towards MG-mediated molecular modification (Hipkiss, 2012). The physiology for the protective response of (CTG) 5 homozygosity recommend that decreased activity of carnosinase stimulates high level of circulating carnosine that provide protection in contrast to hyperglycemia- induced cytotoxic metabolites emerging from oxidative stress and glycation (Freedman et al., 2007). Human serum carnosinase has been suggested as a novel biomarker in (cerebral spinal fluid) CSF (Hu et al., 2007; Perrin et al., 2011).

2.7 Synaptophysin:

Synaptophysin or a major synaptic vesicle protein p38 is expressed in nerve cells and has been marked as a specialized presynaptic marker for neurons (Calhoun et al., 1996). The cortex of the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)- induced monkey model of Parkinson's disease (PD) (Raju et al., 2008), dementia with Lewy bodies and other neurodegenerative disorders are characterized with loss of synaptophysin or low level of synaptophysin (Mukaetova-Ladinska et al., 2013) Dementia is a prevalent problem and trait of many neurodegenerative disorders such as Parkinson's disease (Bossy-Wetzel et al., 2004). While a research study has revealed slight decrease of synaptophysin in the hippocampus of T2DM mice (Carvalho et al., 2015).

2.8 SNAP-25:

Synaptosomal-associated protein of 25 KDa (SNAP-25) is known to be a presynaptic protein necessary for releasing of neurotransmitter (Corradini et al., 2009). The Ser187 of SNAP-25 is phosphorylated by protein kinase C (PKC) to increase the generation of neurotransmitter by recruiting secretory vesicles adjacent to the plasma membrane however a mutant mouse, substituting Ser187 of SNAP-25 with Alanine has been characterized by decreased dopamine release (Kataoka et al., 2011). PD is characterized by continuous degeneration of dopaminergic neurons in the SN pars compacta (Videira & Castro-Caldas, 2018). The level of SNAP-25 was shown low in the cerebral cortex and hippocampus of type 2 diabetic mice model (Carvalho et al., 2015).

2.9 Sorcin:

Sorcin is a calcium sensor protein involved in regulating ER Ca²⁺ by inhibiting ryanodine receptor activity and playing a role in terminating Ca²⁺ induced Ca²⁺ release (Marmugi et al., 2016). The pathogenesis of type 2 diabetes is characterized by dysfunction of β -cells of pancreas, Islets of Langerhans enlarges the mass of β -cell and increases the production of insulin in the promotion of obesity and resistance of insulin (Prentki, 2006). Chronic hyperglycemia increases the necessity of biosynthesis of insulin and also rise the flowing free fatty acids and cytokines lower endoplasmic reticulum (ER) calcium (Ca²⁺) stores (Cunha et al., 2008; Ramadan et al., 2011), causing ER stress and apoptosis (programmed cell death) in prolonged cases (Arruda & Hotamisligil, 2015). Sorcin exhibits a process to dysregulate the function of b-cell in metabolic stress by linking lipotoxicity of b-cell to endoplasmic reticulum calcium and stress of ER Thus, it is revealed that sorcin is expressed lower in pancreatic β -cells under lipotoxic stress, whereas overexpression of sorcin is enough to prevent failure of β -cell and glucose intolerance. Therefore Sorcin may present a target for interference in type 2 Diabetes mellitus (Marmugi et al., 2016). Sorcin has shown to be differentially upregulated in PD (Werner, Heyny-von Haussen, Mall & Wolf, 2008).

2.10 GLP-1 analogues:

The incidence of type 2 diabetes increases the occurrences of Parkinson's disease in type 2 diabetes affected patients. The T2DM and PD share some common pathological mechanisms including dysregulation of insulin. Therefore, the homeostasis of gut-brain axis for healthy central nervous system and peripheral nervous system is an important process in this case (Kim et al., 2017). The type of endogen incretin hormones called Glucagon-like peptide-1 (GLP-1), maintains the releasing of insulin. The outcomes of GLP-1 is dealt by the G-protein and GLP-1 receptor mediate the consequences of GLP-1 which rise the cyclic AMP within cells then protein kinase-A and phosphoinositide 3-kinase are activated resulting phosphorylation of certain subsequent signaling pathways. The pancreas and periphery mostly contain the above mentioned receptors but they are expressed in the frontal cortex, hypothalamus, thalamus, hippocampus, cerebellum and substantia nigra of central nervous system (Athauda

& Foltynie, 2018). These receptors are involved in the generation of nervous tissues, reduction of inflammation, synaptic plasticity, suppressing apoptotic mechanism and boosting the function of mitochondria in the brain (Kim et al., 2017).

Exenatide is synthetically derived from exendin (EX-4). EX-4 is an activator of the GLP-1 hormone regulating insulin and glucose concentration (Aviles-Olmos et al., 2013). EX-4 is applicable in T2DM presently however the therapeutic capability of EX-4 in PD affected persons must be analyzed instantly (Harkavyi et al., 2008). Limiting microglial activation and expression of matrix metalloproteinase-3 by EX-4 has been revealed to influence neural development, support neuronal differentiation, and conserve degeneration of nerve cells through neurotrophic mechanism (Harkavyi et al., 2008; Kim et al., 2009 ; Li et al., 2009; Perry et al., 2002; Perry et al., 2002). Early stage treatment of PD rat model by EX-4, accommodated betterment in behavior and dopamine repairing (Rampersaud et al., 2012). Liraglutide and lixisenatide are the new derivatives of GLP-1 that played a neuroprotective role in PD mouse model by inhibiting apoptosis, by promoting motor impairment and by saving the concentration of tyrosine hydroxylase in the regions of Substantia nigra and basal ganglia (Liu et al., 2015). Another study has concluded that Some of GLP-1 agonists (sitagliptin, a dipeptidyl peptidase-4 inhibitor deactivating GLP-1, and liraglutide, GLP-1 mimetic) enhanced motor functions, prevent nigral degeneration and increase striatal dopamine in rotenone rat model of PD (Badawi et al., 2017). Saxagliptin which is a dipeptidyl peptidase-4 inhibitor has significantly improved motor functions and prevent immunoreaction of tyrosine hydroxylase in Substantia nigra by inhibiting oxidation, inflammation, apoptosis and activating mechanisms of neuroprotection and neurorestoration (Nassar et al., 2015).

2.11 AMPK:

Neurodegeneration in PD may be improved by triggering adenosine monophosphate-activated protein kinase (AMPK) (Choi et al., 2010). Metformin is a class of oral diabetes medication generally used in T2DM and protect against inflammation by activating AMPK (Hang et al., 2015; Ismaiel et al., 2016). Many studies have adverse results showing that neuronal cell loss has been avoided in MPP+/MPTP

animal models of PD by the activation of AMPK (Choi et al., 2010). The result of a research study revealed that AMPK deals decline of dopamine releasing nerve cells in PD mice models and the neurodegeneration in those mice is increased by metformin (Kim et al., 2013). A cohort study among the Taiwanese population surprisingly revealed that the risk of PD was double in the patients suffering from T2DM but this risk can be lowered by using metformin as a conjoint therapy, expressing that metformin is functional to activate AMPK (Wahlqvist et al., 2012). Recently a study has affirmed that prolong application of metformin for type 2 diabetic patients can develop disorders of neurodegeneration like dementia and Parkinson's disease (Kuan et al., 2017).

2.12 Aldehyde dehydrogenase A1 (ADH1A-1):

Aldehyde dehydrogenase A-1 (ALDH1A-1) is a multifaceted enzyme with dehydrogenase, esterase, and anti-oxidant activities and regulates retinoic acid (RA) signaling, which is crucial for the homeostasis of normal brain (Nikhil et al., 2018). ADH1A-1 was differentially overexpressed in the Substantia nigra of PD patient in contrast to controls, which is an enzyme associated with the metabolism of aldehyde (Werner et al., 2008) and dopamine metabolites (Grünblatt & Riederer, 2014). The level of ALDH1A-1 was low in PD brains, and ALDH1A-1 and ALDH2 double knockout mice revealed high HNE (4-hydroxy-2-nonenal) and 3,4-dihydroxyphenylacetaldehyde and remarkable dopaminergic neurodegeneration (Grünblatt & Riederer, 2014). Prolong oxidative stress caused by Cdk5 (Cyclin Dependent Kinase-5) inhibits ALDH1A-1 activity to generate neurotoxicity and overexpression of ALDH1A-1 play vital role in neuroprotection in terms of neurodegenerative disorders (Nikhil et al., 2018). Recently a research study has reported the discovery of an isoform of aldehyde dehydrogenase-1 isoform A3 (ALDH1A-3) as a biomarker of nonfunctional b cells in diabetic mice (Kim-Muller et al., 2016). ALDH1A-3 was markedly absent from normal b cells (Kutlu et al., 2009). A recent study has concluded that ALDH1A-3 is also raised in the pancreatic islets of type 2 diabetic patients (Cinti et al., 2016).

3. DISCUSSION

A link between type 2 diabetes mellitus (T2DM) and Parkinson's disease (PD) has been suggested for decades. Number of common risk factors has been suggested for T2DM and PD, including oxidative stress, autophagy dysregulation, mitochondrial damage (dysfunction), Endoplasmic reticulum stress (malfunction), inflammation and loss of central and peripheral synapse. T2DM and PD both are considered protein conformational diseases. Protein misfolding disorders (PMDs) are diseases in which misfolding, aggregation and accumulation of proteins occur in the disease-specific damaging tissues. Both Parkinson's disease (PD) and type 2 diabetes mellitus (T2DM) are the examples of PMDs (Soto, 2003). The first line of evidence that linked T2DM and PD is protein misfolding and aggregation of amyloid fibres. Amyloid fibres are synthesized from amylin [islet amyloid polypeptide (IAPP)] in T2DM while amyloid like structures are synthesized from α -synuclein in PD (Surguchov, 2016).

Autophagy is a molecular mechanism needed for regulation of cellular physiology and promoting cell survival however defects in autophagy lead to the pathogenesis of several disorders, including Diabetes Mellitus and neurodegeneration. Autophagy eliminates dysfunctional organelles, lipids and miss-folded proteins in T2DM. Additionally, autophagy perform vital role in dysfunction of pancreatic β -cell and resistance to insulin (Yang et al., 2017). Oxidative stress, mitochondrial dysfunction, and protein aggregation play an important role in the pathogenesis of PD; these factors are highly associated to autophagy. The brains of PD patients and animal models of PD have been characterized by dysregulation of autophagy in a recent study, suggesting the developing role of autophagy in PD (Lynch-Day et al., 2012). Mitophagy is a quality control process in healthy neurons for removal of damaged or non-functional mitochondria to prevent cell death (Pickrell & Youle, 2015).

In this review article, it is determined that autophagy induction can be analyzed by different proteins, such as Parkin and PINK 1, Beclin, AMPK, ATG7 and LC3-II in T2DM and PD. Certain studies have concluded that the pathway of PINK1/PARKIN perform vital role in mitochondrial quality control in T2DM and PD (Geisler et al., 2010; Scheele et al., 2007). Dysregulation of autophagy is due to reduction of PINK1/PARKIN level which leads to the pathogenesis of

T2DM and PD. Beclin is a protein associated with the nucleation phase of autophagy, the level of Beclin and Beclin-1 was slightly high in the T2DM and PD respectively suggesting that it minimizes cell death and maximizes the activity of autophagy (Spencer et al. 2009; Carvalho et al., 2015). The level of ATG7 was revealed extremely low in type 2 diabetic mice model, indicating that the elongation phase of the autophagosome synthesis has been agitated (Carvalho et al., 2015). On the other hand, ATG7 is an important autophagy protein needed for degeneration of axonal terminals, and axonopathy linked with neurodegeneration caused by deterioration of axonal autophagy (Komatsu et al., 2007). It has been concluded by a recent study that minimal level of autophagy preserve neurodegeneration (Komatsu et al., 2007). LC3-II is a type of LC3 which is the major constituent of the autophagosomal membrane found both on the inner and outer surface of the membrane. The LC3-II found on the membranes acts as a marker for autophagy (Kabeya et al., 2000; Wu et al., 2006). The level of LC3-II protein has been revealed slightly decreased in the cerebral cortex of type 2 diabetic mice model (Carvalho et al., 2015) indicating that this protein plays vital role in the pathogenesis of T2DM. Autophagy is also important for the pathogenesis of PD however the role of LC3-II in PD related autophagy has not been observed.

Sorcin is another protein dysregulating the function of pancreatic β -cell in metabolic stress by linking lipotoxicity of β -cell to endoplasmic reticulum calcium and stress of ER. It has been revealed that the overexpression of sorcin is enough to prevent failure of β -cells and glucose intolerance. Thus, it may present a target for interference in type 2 diabetes mellitus (Marmugi et al., 2016). Sorcin has shown to be differentially overexpressed in PD (Werner et al., 2008) however the role of sorcin in the development of PD has not been revealed by a study. The above findings suggest that sorcin perform an important role in the pathogenesis of T2DM and PD.

The overexpression of ALDH1A-1 performs vital role in neuroprotection in terms of neurodegeneration (Nikhil et al., 2018). The level of this enzyme has been observed decrease in the SN of PD brain (Werner et al., 2008) indicating its importance in the pathogenesis of PD. The role of ALDH1A-1 in T2DM has not been observed by a study however an isoform of ALDH1 isoform A3

(ALDH1A3) has been discovered as a biomarker of nonfunctional β cells in diabetic mice (Kim-Muller et al., 2016). A recent study has concluded that ALDH1A3 is also raised in the pancreatic islets of type 2 diabetic patients (Cinti et al., 2016).

Synaptophysin and SNAP-25 are presynaptic proteins/markers in neurons for releasing of neurotransmitters. The level of both proteins has been revealed decrease in T2DM and PD (Mukaetova-Ladinska et al., 2013; Carvalho, 2015). Mutation of SNAP-25 protein in mouse model of PD leads to decrease dopamine release (Kataoka et al., 2011). The role of these two proteins in the development of T2DM and PD need further research studies.

Neurodegeneration in PD and inflammation in T2DM may be improved by activating AMPK through the administration of Metformin (Choi et al., 2010; Hang et al., 2015; Ismaiel et al., 2016). Activation of AMPK by Metformin in the treatment of T2DM and PD shows links between these two diseases.

Pancreas contains receptors for GLP1 analogues but they are expressed in different regions of brain (Athauda & Foltynie, 2018). These receptors are also involved in the growth of nervous tissues, prevention of inflammation, synaptic plasticity, suppressing apoptotic process and boosting the function of mitochondria in brain (Kim et al., 2017). GLP-1 may also link the pathogenesis of T2DM and PD. An agonist of GLP-1 known as EX-4 is applicable in the treatment of type 2 diabetic patients however the therapeutic capability of EX-4 in PD affected persons must be trialed instantly (Harkavyi et al., 2008).

The activity of Carnosinase, CNDP2 has been increased in SN of PD suggesting that carnosine make effort for protection against PD (Hipkiss, 2012) and it influences the metabolism of glucose and prevents diabetic deterioration (Lee et al., 2005; Sauerhofer et al., 2007).

4. CONCLUSION

Different literature analyses in this study find out some common proteins involvement in the pathogenesis of both diseases (T2DM and PD). These proteins include amyloid fibres, Parkin & PINK1, Beclin, ATG7, LC3-II, Carnosinase (CNDP2), Synaptophysin, SNAP-25, Sorcin, GLP-1 analogues, AMPK and ADH1A1. Dysregulation of these

proteins causes oxidative stress, autophagy dysregulation, mitochondrial damage (dysfunction), Endoplasmic reticulum stress (malfunction), inflammation and loss of central and peripheral synapses in T2DM and PD.

Although the above findings suggest that type 2 diabetes mellitus and Parkinson's disease are associated through various pathways but there is no conclusive evidence to prove that T2DM is a risk factor for the development of PD. Further prospective, pathological and proteomics studies are required to clarify the correlation of T2DM with PD.

5. ACKNOWLEDGEMENTS

I would like to express my gratitude to my parents especially my mother (late), family and colleagues who supported me throughout my educational career.

Abbreviations used:

T2DM, Type 2 Diabetes Mellitus; PD, Parkinson's Disease; SN, Substantia Nigra; MG, Methylglyoxal; CAR, Carnosine; CNBP1, Carnosine Dipeptidase 1; CNBP2, Carnosine Dipeptidase 2; MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine ; SNAP-25, Synaptosomal-associated protein of 25 KDa; GLP-1, Glucagon-like peptide 1; IAPP, islet amyloid polypeptide; AMPK, Adenosine monophosphate-activated protein kinase; LC3-II, Microtubule-associated protein 1A/1B-light chain 3- II; ADH1A1, Aldehyde dehydrogenase 1A1; ALDH2, Aldehyde Dehydrogenase 2; ALDH1A3, Aldehyde Dehydrogenase 1 Family Member A3; ATG Proteins, Autophagy related proteins; ATG7, Autophagy related 7; ER, Endoplasmic Reticulum; PINK1, PTEN-induced putative kinase 1.

REFERENCES

1. Anglade, P., Vyas, S., Javoy-Agid, F., Herrero, M., Michel, P., Marquez, J., Mouatt-Prigent, A., Ruberg, M., Hirsch, E., & Agid, Y. (1997). Apoptosis and autophagy in nigral neurons of patients with Parkinson's disease. *Histology and histopathology*, *12*(1), 25–31.
2. Arroyo, D., Gaviglio, E., Peralta Ramos, J., Bussi, C., Rodriguez-Galan, M., & Iribarren, P. (2014). Autophagy in inflammation, infection, neurodegeneration and cancer. *International Immunopharmacology*, *18*(1), 55-65. doi: 10.1016/j.intimp.2013.11.001
3. Arruda, A., & Hotamisligil, G. (2015). Calcium Homeostasis and Organelle Function in the Pathogenesis of Obesity and Diabetes. *Cell Metabolism*, *22*(3), 381-397. doi: 10.1016/j.cmet.2015.06.010
4. Athauda, D., & Foltynie, T. (2018). Protective effects of the GLP-1 mimetic exendin-4 in Parkinson's disease. *Neuropharmacology*, *136*, 260-270. doi: 10.1016/j.neuropharm.2017.09.023
5. Aviles-Olmos, I., Dickson, J., Kefalopoulou, Z., Djamshidian, A., Ell, P., Soderlund, T., Whitton, P., Wyse, R., Isaacs, T., Lees, A., Limousin, P., & Foltynie, T. (2013). Exenatide

Sanaullah Khan, Kaleemullah Mandokhail, Habib ur Rehman– **Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease**

- and the treatment of patients with Parkinson's disease. *Journal Of Clinical Investigation*, 123(6), 2730-2736. doi: 10.1172/jci68295
6. Aviles-Olmos, I., Limousin, P., Lees, A., & Toltnie, T. (2012). Parkinson's disease, insulin resistance and novel agents of neuroprotection. *Brain*, 136(2), 374-384. doi: 10.1093/brain/aws009
 7. Badawi, G., Abd El Fattah, M., Zaki, H., & El Sayed, M. (2017). Sitagliptin and liraglutide reversed nigrostriatal degeneration of rodent brain in rotenone-induced Parkinson's disease. *Inflammopharmacology*, 25(3), 369-382. doi: 10.1007/s10787-017-0331-6
 8. Bai, H., Inoue, J., Kawano, T., & Inazawa, J. (2012). A transcriptional variant of the LC3A gene is involved in autophagy and frequently inactivated in human cancers. *Oncogene*, 31(40), 4397-4408. doi: 10.1038/ncr.2011.613.
 9. Banerjee, R., Beal, M., & Thomas, B. (2010). Autophagy in neurodegenerative disorders: pathogenic roles and therapeutic implications. *Trends In Neurosciences*, 33(12), 541-549. doi: 10.1016/j.tins.2010.09.001
 10. Bellia, F., Vecchio, G., & Rizzarelli, E. (2014). Carnosinases, Their Substrates and Diseases. *Molecules*, 19(2), 2299-2329. doi: 10.3390/molecules19022299
 11. Bossy-Wetzell, E., Schwarzenbacher, R., & Lipton, S. (2004). Molecular pathways to neurodegeneration. *Nature Medicine*, 10(S7), S2-S9. doi: 10.1038/nm1067
 12. Butler, A., Janson, J., Bonner-Weir, S., Ritzel, R., Rizza, R., & Butler, P. (2003). -Cell Deficit and Increased -Cell Apoptosis in Humans With Type 2 Diabetes. *Diabetes*, 52(1), 102-110. doi: 10.2337/diabetes.52.1.102
 13. Butterworth, R., Wassif, W., Sherwood, R., Gerges, A., Poyser, K., Garthwaite, J., Peters, T., & Bath, P. (1996). Serum Neuron-Specific Enolase, Carnosinase, and Their Ratio in Acute Stroke. *Stroke*, 27(11), 2064-2068. doi: 10.1161/01.str.27.11.2064
 14. Calhoun, M., Jucker, M., Martin, L., Thinakaran, G., Price, D., & Mouton, P. (1996). Comparative evaluation of synaptophysin-based methods for quantification of synapses. *Journal Of Neurocytology*, 25(1), 821-828. doi: 10.1007/bf02284844
 15. Carvalho, C., Santos, M., Oliveira, C., & Moreira, P. (2015). Alzheimer's disease and type 2 diabetes-related alterations in brain mitochondria, autophagy and synaptic markers. *Biochimica Et Biophysica Acta (BBA) - Molecular Basis Of Disease*, 1852(8), 1665-1675. doi: 10.1016/j.bbdis.2015.05.001
 16. Cataldo, A., Hamilton, D., Barnett, J., Paskevich, P., & Nixon, R. (1996). Properties of the endosomal-lysosomal system in the human central nervous system: disturbances mark most neurons in populations at risk to degenerate in Alzheimer's disease. *The Journal of Neuroscience*, 16(1), 186-199. doi: 10.1523/jneurosci.16-01-00186.1996
 17. Chao, Y., Chew, L., Deng, X., & Tan, E. (2015). Nonmotor symptoms in sporadic versus familial forms of Parkinson's disease. *Neurodegenerative Disease Management*, 5(2), 147-153. doi: 10.2217/nmt.14.57
 18. Chen, L., Magliano, D., & Zimmet, P. (2011). The worldwide epidemiology of type 2 diabetes mellitus—present and future perspectives. *Nature Reviews Endocrinology*, 8(4), 228-236. doi: 10.1038/nrendo.2011.183
 19. Choi, J., Park, C., & Jeong, J. (2010). AMP-activated protein kinase is activated in Parkinson's disease models mediated by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine. *Biochemical And Biophysical Research Communications*, 391(1), 147-151. doi: 10.1016/j.bbrc.2009.11.022
 20. Cinti, F., Bouchi, R., Kim-Muller, J., Ohmura, Y., Sandoval, P., Masini, M., Marselli, L., Suleiman, M., Ratner, L., Marchetti, P., & Accili, D. (2016). Evidence of β -Cell Dedifferentiation in Human Type 2 Diabetes. *The Journal Of Clinical Endocrinology & Metabolism*, 101(3), 1044-1054. doi: 10.1210/jc.2015-2860
 21. Coleman, M. (2002). Axon pathology in neurological disease: a neglected therapeutic target. *Trends In Neurosciences*, 25(10), 532-537. doi: 10.1016/s0166-2236(02)02255-5

Sanaullah Khan, Kaleemullah Mandokhail, Habib ur Rehman– **Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease**

22. Corradini, I., Verderio, C., Sala, M., Wilson, M., & Matteoli, M. (2009). SNAP-25 in Neuropsychiatric Disorders. *Annals Of The New York Academy Of Sciences*, 1152(1), 93-99. doi: 10.1111/j.1749-6632.2008.03995.x
23. Cunha, D. A., Hekerman, P., Ladriere, L., Bazarra-Castro, A., Ortis, F., Wakeham, M. C., Moore, F., Rasschaert, J., Cardozo, A., Bellomo, E., Overbergh, L., Mathieu, C., Lupi, R., Hai, T., Herchuelz, A., Marchetti, P., Rutter, G., Eizirik, D., & Cnop, M. (2008). Initiation and execution of lipotoxic ER stress in pancreatic β -cells. *Journal Of Cell Science*, 121(14), 2308-2318. doi: 10.1242/jcs.026062
24. Dukic-Stefanovic, S., Schinzel, R., Riederer, P., & Münch*, G. (2001). Journal search results - Cite This For Me. *Biogerontology*, 2(1), 19-34. doi: 10.1023/a:1010052800347
25. Edinburg Regional Medical Center (ER), Parkinson's disease. (April 16, 2015). Available at: <http://www.edinburgregional.com/hospitalservices/neurosurgery/parkinsonsdisease#V.TvILtKqqko>
26. Fatima, S., Haque, R., Jadya, P., Shamsuzzama, Kumar, L., & Nazir, A. (2014). Ida-1, the *Caenorhabditis elegans* Orthologue of Mammalian Diabetes Autoantigen IA-2, Potentially Acts as a Common Modulator between Parkinson's Disease and Diabetes: Role of Daf-2/Daf-16 Insulin Like Signalling Pathway. *Plos ONE*, 9(12), e113986. doi: 10.1371/journal.pone.0113986
27. Fontana, M., Pinnen, F., Lucente, G., & Pecci, L. (2002). Prevention of peroxynitrite-dependent damage by carnosine and related sulphonamido pseudodipeptides. *Cellular And Molecular Life Sciences (CMLS)*, 59(3), 546-551. doi: 10.1007/s00018-002-8446-2
28. Freedman, B., Hicks, P., Sale, M., Pierson, E., Langefeld, C., Rich, S., Xu, J., McDonough, C., Janssen, B., Yard, B., van der Woude, F., & Bowden, D. (2007). A leucine repeat in the carnosinase gene CNDP1 is associated with diabetic end-stage renal disease in European Americans. *Nephrology Dialysis Transplantation*, 22(4), 1131-1135. doi: 10.1093/ndt/gfl717
29. Fujioka, K. (2007). Pathophysiology of Type 2 Diabetes and the Role of Incretin Hormones and Beta-Cell Dysfunction. *Journal Of The American Academy Of Physician Assistants*, 20(12), 3-8. doi: 10.1097/01720610-200712000-00001
30. Geisler, S., Holmström, K. M., Treis, A., Skujat, D., Weber, S., Fiesel, F., Kahle, P., & Springer, W. (2010). The PINK1/Parkin-mediated mitophagy is compromised by PD-associated mutations. *Autophagy*, 6(7), 871-878. doi: 10.4161/auto.6.7.13286
31. Genetic basis of type 1 and type 2 diabetes, obesity, and their complications. Advances and emerging opportunities in diabetes research: a Strategic Planning report of the DMICC. (2011). www2.niddk.nih.gov/NR.
32. Grünblatt, E., & Riederer, P. (2014). Aldehyde dehydrogenase (ALDH) in Alzheimer's and Parkinson's disease. *Journal Of Neural Transmission*, 123(2), 83-90. doi: 10.1007/s00702-014-1320-1
33. Hang, L., Thundiyil, J., & Lim, K. (2015). Mitochondrial dysfunction and Parkinson disease: a Parkin-AMPK alliance in neuroprotection. *Annals Of The New York Academy Of Sciences*, 1350(1), 37-47. doi: 10.1111/nyas.12820
34. Harkavyi, A., Abuirmeileh, A., Lever, R., Kingsbury, A., Biggs, C., & Whitton, P. (2008). Glucagon-like peptide 1 receptor stimulation by exendin-4 reverses key deficits in distinct rodent models of Parkinson's disease. *Journal Of Neuroinflammation*, 5(1), 19. doi: 10.1186/1742-2094-5-19
35. He, H., Dang, Y., Dai, F., Guo, Z., Wu, J., She, X., Pei, Y., Chen, Y., Ling, W., Wu, C., Zhao, S., Liu, J., & Yu, L. (2003). Post-translational Modifications of Three Members of the Human MAP1LC3 Family and Detection of a Novel Type of Modification for MAP1LC3B. *Journal Of Biological Chemistry*, 278(31), 29278-29287. doi: 10.1074/jbc.m303800200
36. Hipkiss, A. (2005). Glycation, ageing and carnosine: Are carnivorous diets beneficial?. *Mechanisms Of Ageing And Development*, 126(10), 1034-1039. doi: 10.1016/j.mad.2005.05.002

Sanaullah Khan, Kaleemullah Mandokhail, Habib ur Rehman– **Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease**

37. Hipkiss, A. (2007). Could Carnosine or Related Structures Suppress Alzheimer's Disease?. *Journal Of Alzheimer's Disease*, *11*(2), 229-240. doi: 10.3233/jad-2007-11210
38. Hoshino, A., Ariyoshi, M., Okawa, Y., Kaimoto, S., Uchihashi, M., Fukai, K., Iwai-Kanai, E., Ikeda, K., Ueyama, T., Ogata, T., & Matoba, S. (2014). Inhibition of p53 preserves Parkin-mediated mitophagy and pancreatic β -cell function in diabetes. *Proceedings Of The National Academy Of Sciences*, *111*(8), 3116-3121. doi: 10.1073/pnas.1318951111
39. Hu, F., Manson, J., Stampfer, M., Colditz, G., Liu, S., Solomon, C., & Willett, W. (2001). Diet, Lifestyle, and the Risk of Type 2 Diabetes Mellitus in Women. *New England Journal Of Medicine*, *345*(11), 790-797. doi: 10.1056/nejmoa010492
40. Hu, Y., Hosseini, A., Kauwe, J., Gross, J., Cairns, N., Goate, A., Fagan, A., Townsend, R., & Holtzman, D. (2007). Identification and validation of novel CSF biomarkers for early stages of Alzheimer's disease. *PROTEOMICS – CLINICAL APPLICATIONS*, *1*(11), 1373-1384. doi: 10.1002/prca.200600999
41. Huang, Y., Duan, J., Chen, H., Chen, M., & Chen, G. (2005). Separation and determination of carnosine-related peptides using capillary electrophoresis with laser-induced fluorescence detection. *ELECTROPHORESIS*, *26*(3), 593-599. doi: 10.1002/elps.200406130
42. Ichimura, Y., Kirisako, T., Takao, T., Satomi, Y., Shimonishi, Y., Ishihara, N., Mizushima, N., Tanida, I., Kominami, E., Ohsumi, M., Noda, T., & Ohsumi, Y. (2000). A ubiquitin-like system mediates protein lipidation. *Nature*, *408*(6811), 488-492. doi: 10.1038/35044114
43. Imtiaz, N., Mehreen, S., Saeed, K., Akhtar, N., Ur, H., Rehman, Amin, S., U, A, Rehman, Ali, J., Ayub, M., & Bibi, Z. (2016). Study of prevalence of Parkinson's disease in elderly population in Rawalpindi, Pakistan. *Journal of Entomology and Zoology Studies*, *4*(6), 845-847.
44. Ismaiel, A., Espinosa-Oliva, A., Santiago, M., García-Quintanilla, A., Oliva-Martín, M., Herrera, A., Venero, J., & de Pablos, R. (2016). Metformin, besides exhibiting strong in vivo anti-inflammatory properties, increases mptp-induced damage to the nigrostriatal dopaminergic system. *Toxicology And Applied Pharmacology*, *298*, 19-30. doi: 10.1016/j.taap.2016.03.004
45. Janssen, B., Hohenadel, D., Brinkkoetter, P., Peters, V., Rind, N., Fischer, C., Rychlik, I., Cerna, M., Romzova, M., de Heer, E., Baelde, H., Bakker, S., Zirje, M., Rondeau, E., Mathieson, P., Saleem, M., Meyer, J., Koppel, H., Sauerhoefer, S., Bartram, C., Nawroth, P., Hammes, H., Yard, B., Zschocke, J., & van der Woude, F. (2005). Carnosine as a Protective Factor in Diabetic Nephropathy: Association With a Leucine Repeat of the Carnosinase Gene CNDP1. *Diabetes*, *54*(8), 2320-2327. doi: 10.2337/diabetes.54.8.2320
46. Kabeya, Y., Mizushima, N., Ueno, T., Yamamoto, A., Kirisako, T., Noda, T., Kominami, E., Ohsumi, Y., & Yoshimori, T. (2000). LC3, a mammalian homologue of yeast Agg8p, is localized in autophagosome membranes after processing. *The EMBO journal*, *19*(21), 5720–5728. doi: 10.1093/emboj/19.21.5720
47. Kahn, S. (2003). The relative contributions of insulin resistance and beta-cell dysfunction to the pathophysiology of Type 2 diabetes. *Diabetologia*, *46*(1), 3-19. doi: 10.1007/s00125-002-1009-0
48. Kalia, L., & Lang, A. (2015). Parkinson's disease. *The Lancet*, *386*(9996), 896-912. doi: 10.1016/s0140-6736(14)61393-3
49. Kataoka, M., Yamamori, S., Suzuki, E., Watanabe, S., Sato, T., Miyaoka, H., Azuma, S., Ikegami, S., Kuwahara, R., Suzuki-Migishima, R., Nakahara, Y., Nihonmatsu, I., Inokuchi, K., Katoh-Fukui, Y., Yokoyama, M., & Takahashi, M. (2011). A Single Amino Acid Mutation in SNAP-25 Induces Anxiety-Related Behavior in Mouse. *Plos ONE*, *6*(9), e25158. doi: 10.1371/journal.pone.0025158
50. Kim, D. S., Choi, H. I., Wang, Y., Luo, Y., Hoffer, B., & Greig, N. (2017). A new treatment strategy for Parkinson's disease through the gut–brain axis: the glucagon-like peptide-1 receptor pathway. *Cell transplantation*, *26*(9), 1560-1571.

Sanaullah Khan, Kaleemullah Mandokhail, Habib ur Rehman– **Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease**

51. Kim, S., Moon, M., & Park, S. (2009). Exendin-4 protects dopaminergic neurons by inhibition of microglial activation and matrix metalloproteinase-3 expression in an animal model of Parkinson's disease. *Journal Of Endocrinology*, 202(3), 431-439. doi: 10.1677/joe-09-0132
52. Kim, T., Cho, H., Choi, S., Suguira, Y., Hayasaka, T., Setou, M., Koh, H., Mi Hwang, E., Park, J., Kang, S., Kim, H., Kim, H., & Sun, W. (2013). (ADP-ribose) polymerase 1 and AMP-activated protein kinase mediate progressive dopaminergic neuronal degeneration in a mouse model of Parkinson's disease. *Cell Death & Disease*, 4(11), e919-e919. doi: 10.1038/cddis.2013.447
53. Kim-Muller, J., Fan, J., Kim, Y., Lee, S., Ishida, E., Blaner, W., & Accili, D. (2016). Aldehyde dehydrogenase 1a3 defines a subset of failing pancreatic β cells in diabetic mice. *Nature Communications*, 7(1). doi: 10.1038/ncomms12631
54. Klionsky, D. (2000). Autophagy as a Regulated Pathway of Cellular Degradation. *Science*, 290(5497), 1717-1721. doi: 10.1126/science.290.5497.1717
55. Komatsu, M., Wang, Q., Holstein, G., Friedrich, V., Iwata, J., Kominami, E., Chait, B., Tanaka, K., & Yue, Z. (2007). Essential role for autophagy protein Atg7 in the maintenance of axonal homeostasis and the prevention of axonal degeneration. *Proceedings Of The National Academy Of Sciences*, 104(36), 14489-14494. doi: 10.1073/pnas.0701311104
56. Kuan, Y., Huang, K., Lin, C., Hu, C., & Kao, C. (2017). Effects of metformin exposure on neurodegenerative diseases in elderly patients with type 2 diabetes mellitus. *Progress In Neuro-Psychopharmacology And Biological Psychiatry*, 79, 77-83. doi: 10.1016/j.pnpbp.2017.06.002
57. Kumar, K., Debjit, B., Chandira, M., Pankaj, T., & Shideshwar, S. (2010). Role of community pharmacist care and management of Parkinsonism disease. *Journal of Chemical and Pharmaceutical Research*, 2(1), 315-318.
58. Kutlu, B., Burdick, D., Baxter, D., Rasschaert, J., Flamez, D., Eizirik, D., Welsh, N., Goodman, N., & Hood, L. (2009). Detailed transcriptome atlas of the pancreatic beta cell. *BMC Medical Genomics*, 2(1). doi: 10.1186/1755-8794-2-3
59. La Mendola, D., Sortino, S., Vecchio, G., & Rizzarelli, E. (2002). Synthesis of New Carnosine Derivatives of β -Cyclodextrin and Their Hydroxyl Radical Scavenger Ability. *Helvetica Chimica Acta*, 85(6), 1633-1643. doi: 10.1002/1522-2675(200206)85:6<1633::aid-hlca1633>3.0.co;2-g
60. Lee, Y., Hsu, C., Lin, M., Liu, K., & Yin, M. (2005). Histidine and carnosine delay diabetic deterioration in mice and protect human low density lipoprotein against oxidation and glycation. *European Journal Of Pharmacology*, 513(1-2), 145-150. doi: 10.1016/j.ejphar.2005.02.010
61. Li, H., Li, S., Yu, Z., Shelbourne, P., & Li, X. (2001). Huntingtin Aggregate-Associated Axonal Degeneration is an Early Pathological Event in Huntington's Disease Mice. *The Journal Of Neuroscience*, 21(21), 8473-8481. doi: 10.1523/jneurosci.21-21-08473.2001
62. Li, Y., Perry, T., Kindy, M., Harvey, B., Tweedie, D., Holloway, H., Powers, K., Shen, H., Egan, J., Sambamurti, K., Brossi, A., Lahiri, D., Mattson, M., Hoffer, B., Wang, Y., & Greig, N. (2009). GLP-1 receptor stimulation preserves primary cortical and dopaminergic neurons in cellular and rodent models of stroke and Parkinsonism. *Proceedings Of The National Academy Of Sciences*, 106(4), 1285-1290. doi: 10.1073/pnas.0806720106
63. Licker, V., Côte, M., Lobrinus, J., Rodrigo, N., Kövari, E., Hochstrasser, D., Turck, N., Sanchez, J., & Burkhard, P. (2012). Proteomic profiling of the substantia nigra demonstrates CNDP2 overexpression in Parkinson's disease. *Journal Of Proteomics*, 75(15), 4656-4667. doi: 10.1016/j.jprot.2012.02.032
64. Lima, M., Targa, A., Nosedà, A., Rodrigues, L., Delattre, A., Santos, F., Fortes, M., Maturana, M., & C. Ferraz, A. (2014). Does Parkinson's Disease and Type-2 Diabetes Mellitus Present Common Pathophysiological Mechanisms and Treatments?. *CNS &*

Sanaullah Khan, Kaleemullah Mandokhail, Habib ur Rehman– **Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease**

- Neurological Disorders - Drug Targets*, 13(3), 418-428. doi: 10.2174/18715273113126660155
65. Lin, W., Lewis, J., Yen, S., Hutton, M., & Dickson, D. (2003). Ultrastructural neuronal pathology in transgenic mice expressing mutant (P301L) human tau. *Journal Of Neurocytology*, 32(9), 1091-1105. doi: 10.1023/b:neur.0000021904.61387.95
 66. Liu, W., Jalewa, J., Sharma, M., Li, G., Li, L., & Hölscher, C. (2015). Neuroprotective effects of lixisenatide and liraglutide in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. *Neuroscience*, 303, 42-50. doi: 10.1016/j.neuroscience.2015.06.054
 67. Lucin, K., O'Brien, C., Bieri, G., Czirr, E., Moshier, K., Abbey, R., Mastroeni, D., Rogers, J., Spencer, B., Masliah, E., & Wyss-Coray, T. (2013). Microglial Beclin 1 Regulates Retromer Trafficking and Phagocytosis and Is Impaired in Alzheimer's Disease. *Neuron*, 79(5), 873-886. doi: 10.1016/j.neuron.2013.06.046
 68. Lynch-Day, M., Mao, K., Wang, K., Zhao, M., & Klionsky, D. (2012). The Role of Autophagy in Parkinson's Disease. *Cold Spring Harbor Perspectives In Medicine*, 2(4), a009357-a009357. doi: 10.1101/cshperspect.a009357
 69. Lynch-Day, M., Mao, K., Wang, K., Zhao, M., & Klionsky, D. (2012). The Role of Autophagy in Parkinson's Disease. *Cold Spring Harbor Perspectives In Medicine*, 2(4), a009357-a009357. doi: 10.1101/cshperspect.a009357
 70. Maitra A, Abbas AK (2005) The endocrine system. In: Kumar V, Abbas AK, Fausto N, editors. *Pathologic Basis of Disease*. Philadelphia: Elsevier Saunders. pp. 1155–1226
 71. Marmugi, A., Parnis, J., Chen, X., Carmichael, L., Hardy, J., Mannan, N., Marchetti, P., Piemonti, L., Bosco, D., Johnson, P., Shapiro, J., Cruciani-Guglielmacci, C., Magnan, C., Ibberson, M., Thorens, B., Valdivia, H., Rutter, G., & Leclerc, I. (2016). Sorcin Links Pancreatic β -Cell Lipotoxicity to ER Ca^{2+} Stores. *Diabetes*, 65(4), 1009-1021. doi: 10.2337/db15-1334
 72. Marrif, H., & Al-Sunousi, S. (2016). Pancreatic β Cell Mass Death. *Frontiers In Pharmacology*, 7(83), 1-16. doi: 10.3389/fphar.2016.00083
 73. Mukaetova-Ladinska, E., Andras, A., Milne, J., Abdel-All, Z., Borr, I., Jaros, E., Perry, R., Honer, W., Cleghorn, A., Doherty, J., McIntosh, G., Perry, E., Kalaria, R., & McKeith, I. (2013). Synaptic Proteins and Choline Acetyltransferase Loss in Visual Cortex in Dementia With Lewy Bodies. *Journal Of Neuropathology & Experimental Neurology*, 72(1), 53-60. doi: 10.1097/nen.0b013e31827c5710
 74. Nascimento-Ferreira, I., Nóbrega, C., Vasconcelos-Ferreira, A., Onofre, I., Albuquerque, D., Avelaira, C., Hirai, H., Déglon, N., & Pereira de Almeida, L. (2013). Beclin 1 mitigates motor and neuropathological deficits in genetic mouse models of Machado–Joseph disease. *Brain*, 136(7), 2173-2188. doi: 10.1093/brain/awt144
 75. Nassar, N., Al-Shorbagy, M., Arab, H., & Abdallah, D. (2015). Saxagliptin: A novel antiparkinsonian approach. *Neuropharmacology*, 89, 308-317. doi: 10.1016/j.neuropharm.2014.10.007
 76. Nikhil, K., Viccaro, K., & Shah, K. (2018). Multifaceted Regulation of ALDH1A1 by Cdk5 in Alzheimer's Disease Pathogenesis. *Molecular Neurobiology*, 56(2), 1366-1390. doi: 10.1007/s12035-018-1114-9
 77. Nixon, R., Wegiel, J., Kumar, A., Yu, W., Peterhoff, C., Cataldo, A., & Cuervo, A. (2005). Extensive Involvement of Autophagy in Alzheimer Disease: An Immuno-Electron Microscopy Study. *Journal Of Neuropathology & Experimental Neurology*, 64(2), 113-122. doi: 10.1093/jnen/64.2.113
 78. Parkinson's Disease Information. (2015). What is parkinson's disease? (Updated: June 23, 2009) Available at: <http://www.parkinsons.org/faq.html>.
 79. Perrin, R., Craig-Schapiro, R., Malone, J., Shah, A., Gilmore, P., Davis, A., Roe, C., Peskind, E., Li, G., Galasko, D., Clark, C., Quinn, J., Kaye, J., Morris, J., Holtzman, D., Townsend, R., & Fagan, A. (2011). Identification and Validation of Novel Cerebrospinal Fluid Biomarkers for Staging Early Alzheimer's Disease. *Plos ONE*, 6(1), e16032. doi: 10.1371/journal.pone.0016032

Sanauallah Khan, Kaleemullah Mandokhail, Habib ur Rehman– **Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease**

80. Perry, T., Haughey, N., Mattson, M., Egan, J., & Greig, N. (2002). Protection and Reversal of Excitotoxic Neuronal Damage by Glucagon-Like Peptide-1 and Exendin-4. *Journal Of Pharmacology And Experimental Therapeutics*, 302(3), 881-888. doi: 10.1124/jpet.102.037481
81. Perry, T., Lahiri, D., Chen, D., Zhou, J., Shaw, K., Egan, J., & Greig, N. (2002). A Novel Neurotrophic Property of Glucagon-Like Peptide I: A Promoter of Nerve Growth Factor-Mediated Differentiation in PC12 Cells. *Journal Of Pharmacology And Experimental Therapeutics*, 300(3), 958-966. doi: 10.1124/jpet.300.3.958
82. Pickford, F., Masliah, E., Britschgi, M., Lucin, K., Narasimhan, R., Jaeger, P., Small, S., Spencer, B., Rockenstein, E., Levine, B., & Wyss-Coray, T. (2008). The autophagy-related protein beclin 1 shows reduced expression in early Alzheimer disease and regulates amyloid β accumulation in mice. *Journal Of Clinical Investigation*, 118(6), 2190-2199. doi: 10.1172/jci33585
83. Pickrell, A., & Youle, R. (2015). The Roles of PINK1, Parkin, and Mitochondrial Fidelity in Parkinson's Disease. *Neuron*, 85(2), 257-273. doi: 10.1016/j.neuron.2014.12.007
84. Pratley, R., & Weyer, C. (2001). The role of impaired early insulin secretion in the pathogenesis of Type II diabetes mellitus. *Diabetologia*, 44(8), 929-945. doi: 10.1007/s001250100580
85. Prentki, M. (2006). Islet cell failure in type 2 diabetes. *Journal Of Clinical Investigation*, 116(7), 1802-1812. doi: 10.1172/jci29103
86. Pressley, J., Louis, E., Tang, M., Cote, L., Cohen, P., Glied, S., & Mayeux, R. (2003). The impact of comorbid disease and injuries on resource use and expenditures in parkinsonism. *Neurology*, 60(1), 87-93. doi: 10.1212/wnl.60.1.87
87. Pubill, D., Verdaguer, E., Sureda, F., Camins, A., Pallàs, M., Camarasa, J., & Escubedo, E. (2002). Carnosine prevents methamphetamine-induced gliosis but not dopamine terminal loss in rats. *European Journal Of Pharmacology*, 448(2-3), 165-168. doi: 10.1016/s0014-2999(02)01949-0
88. R. Hipkiss, A. (2012). Parkinson's Disease and Type-2 Diabetes: Methylglyoxal may be a Common Causal Agent; Carnosine could be Protective. *Molecular Medicine & Therapeutics*, 01(02). doi: 10.4172/2324-8769.1000104
89. R. Hipkiss, A. (2012). Parkinson's Disease and Type-2 Diabetes: Methylglyoxal may be a Common Causal Agent; Carnosine could be Protective. *Molecular Medicine & Therapeutics*, 01(02). doi: 10.4172/2324-8769.1000104
90. Raju, D., Ahern, T., Shah, D., Wright, T., Standaert, D., Hall, R., & Smith, Y. (2008). Differential synaptic plasticity of the corticostriatal and thalamostriatal systems in an MPTP-treated monkey model of parkinsonism. *European Journal Of Neuroscience*, 27(7), 1647-1658. doi: 10.1111/j.1460-9568.2008.06136.x
91. Ramadan, J., Steiner, S., O'Neill, C., & Nunemaker, C. (2011). The central role of calcium in the effects of cytokines on beta-cell function: Implications for type 1 and type 2 diabetes. *Cell Calcium*, 50(6), 481-490. doi: 10.1016/j.ceca.2011.08.005
92. Rampersaud, N., Harkavyi, A., Giordano, G., Lever, R., Whitton, J., & Whitton, P. (2012). Retracted: Exendin-4 reverts behavioural and neurochemical dysfunction in a pre-motor rodent model of Parkinson's disease with noradrenergic deficit. *British Journal Of Pharmacology*, 167(7), 1467-1479. doi: 10.1111/j.1476-5381.2012.02100.x
93. Rewar, S. (2015). A systematic review on Parkinson's disease (PD). *Indian Journal of Research in Pharmacy and Biotechnology*, 3(2), 176.
94. Saeedi, P., Petersohn, I., Salpea, P., Malanda, B., Karuranga, S., Unwin, N., Colagiuri, S., Guariguata, L., Motala, A., Ogurtsova, K., Shaw, J., Bright, D., & Williams, R. (2019). Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas, 9th edition. *Diabetes Research And Clinical Practice*, 157, 107843. doi: 10.1016/j.diabres.2019.107843
95. Sauerhofer, S., Yuan, G., Braun, G., Deinzer, M., Neumaier, M., Gretz, N., Floege, J., Kriz, W., van der Woude, F., & Moeller, M. (2007). L-Carnosine, a Substrate of

Sanallah Khan, Kaleemullah Mandokhail, Habib ur Rehman– **Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease**

- Carnosinase-1, Influences Glucose Metabolism. *Diabetes*, 56(10), 2425-2432. doi: 10.2337/db07-0177
96. Scheele, C., Nielsen, A., Walden, T., Sewell, D., Fischer, C., Brogan, R., Petrovic, N., Larsson, O., Tesch, P., Wennmalm, K., Hutchinson, D., Cannon, B., Wahlestedt, C., Pedersen, B., & Timmons, J. (2007). Altered regulation of the PINK1 locus: a link between type 2 diabetes and neurodegeneration?. *The FASEB Journal*, 21(13), 3653-3665. doi: 10.1096/fj.07-8520com
97. Shafique, H., Blagrove, A., Chung, A., & Logendrarajah, R. (2011). Causes of Parkinson's disease: Literature Review. *Journal Of Parkinsonism And Restless Legs Syndrome*, 1(1), 5-7. doi: 10.7157/jprls.2011v1n1pp5-7
98. Soto, C. (2003). Unfolding the role of protein misfolding in neurodegenerative diseases. *Nature Reviews Neuroscience*, 4(1), 49-60. doi: 10.1038/nrn1007
99. Spencer, B., Potkar, R., Trejo, M., Rockenstein, E., Patrick, C., Gindi, R., Adame, A., Wyss-Coray, T., & Masliah, E. (2009). Beclin 1 Gene Transfer Activates Autophagy and Ameliorates the Neurodegenerative Pathology in -Synuclein Models of Parkinson's and Lewy Body Diseases. *Journal Of Neuroscience*, 29(43), 13578-13588. doi: 10.1523/jneurosci.4390-09.2009
100. Stumvoll, M., Goldstein, B., & van Haeften, T. (2005). Type 2 diabetes: principles of pathogenesis and therapy. *The Lancet*, 365(9467), 1333-1346. doi: 10.1016/s0140-6736(05)61032-x
101. Surguchov, A. (2016). Association between Type-2 diabetes and Parkinson's disease: a cross-talk between amylin and α -synuclein. *Journal Of Diabetes Mellitus And Metabolic Syndrome*, 1(1), 1-7. doi: 10.28967/jdmms.2016.01.16001
102. Tang, S., Arumugam, T., Cutler, R., Jo, D., Magnus, T., Chan, S., Mughal, M., Telljohann, R., Nassar, M., Ouyang, X., Calderan, A., Ruzza, P., Guiotto, A., & Mattson, M. (2007). Neuroprotective actions of a histidine analogue in models of ischemic stroke. *Journal Of Neurochemistry*, 101(3), 729-736. doi: 10.1111/j.1471-4159.2006.04412.x
103. Tian, T., Li, Z., & Lu, H. (2015). Common pathophysiology affecting diabetic retinopathy and Parkinson's disease. *Medical Hypotheses*, 85(4), 397-398. doi: 10.1016/j.mehy.2015.06.016
104. Trombley, P., Horning, M., & Blakemore, L. (2000). Interactions between carnosine and zinc and copper: implications for neuromodulation and neuroprotection. *BIOCHEMISTRY C/C OF BIOKHMIIA*, 65(7), 807-816.
105. Videira, P., & Castro-Caldas, M. (2018). Linking Glycation and Glycosylation With Inflammation and Mitochondrial Dysfunction in Parkinson's Disease. *Frontiers In Neuroscience*, 12. doi: 10.3389/fnins.2018.00381
106. Wahlqvist, M., Lee, M., Hsu, C., Chuang, S., Lee, J., & Tsai, H. (2012). Metformin-inclusive sulfonylurea therapy reduces the risk of Parkinson's disease occurring with Type 2 diabetes in a Taiwanese population cohort. *Parkinsonism & related disorders*, 18(6), 753-758. doi:10.1016/j.parkreldis.2012.03.010
107. Werner, C., Heyny-von Haussen, R., Mall, G., & Wolf, S. (2008). Proteome analysis of human substantia nigra in Parkinson's disease. *Proteome Science*, 6(1), 8. doi: 10.1186/1477-5956-6-8
108. World Health Organization (WHO). Diabetes fact sheet 312. January 2011. Available at: <http://www.who.int/mediacentre/factsheets/fs312/en/>. Accessed May 8, 2013.
109. Wu, J., Dang, Y., Su, W., Liu, C., Ma, H., Shan, Y., Pei, Y., Wan, B., Guo, J., & Yu, L. (2006). Molecular cloning and characterization of rat LC3A and LC3B—Two novel markers of autophagosome. *Biochemical And Biophysical Research Communications*, 339(1), 437-442. doi: 10.1016/j.bbrc.2005.10.211
110. Wu, Y., Ding, Y., Tanaka, Y., & Zhang, W. (2014). Risk Factors Contributing to Type 2 Diabetes and Recent Advances in the Treatment and Prevention. *International Journal Of Medical Sciences*, 11(11), 1185-1200. doi: 10.7150/ijms.10001

Sanaullah Khan, Kaleemullah Mandokhail, Habib ur Rehman– **Determination of Proteomics Correlation between Type 2 Diabetes Mellitus and Parkinson's disease**

111. Xu, Q., Park, Y., Huang, X., Hollenbeck, A., Blair, A., Schatzkin, A., & Chen, H. (2011). Diabetes and Risk of Parkinson's Disease. *Diabetes Care*, *34*(4), 910-915. doi: 10.2337/dc10-1922
112. Yang, J., Lu, C., Kuo, S., Hsu, Y., Tsai, S., Chen, S., Chen, Y., Lin, Y., Huang, Y., Chen, C., Lin, W., Liao, W., Lin, W., Liu, Y., Sheu, J., & Tsai, F. (2017). Autophagy and its link to type II diabetes mellitus. *Biomedicine*, *7*(2), 1-12. doi: 10.1051/bmdcn/2017070201
113. Yu, W., Cuervo, A., Kumar, A., Peterhoff, C., Schmidt, S., Lee, J., Mohan, P., Mercken, M., Farmery, M., Tjernberg, L., Jiang, Y., Duff, K., Uchiyama, Y., Našlund, J., Mathews, P., Cataldo, A., & Nixon, R. (2005). Macroautophagy- a novel β -amyloid peptide-generating pathway activated in Alzheimer's disease. *Journal of Cell Biology*, *171*(1), 87-98. doi: 10.1083/jcb.200505082